

Gene Expression Analysis : Tamoxifen Resistance in Breast Cancer

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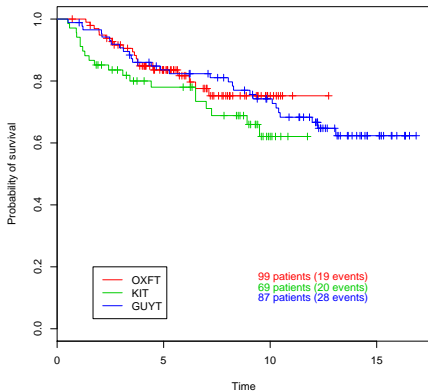
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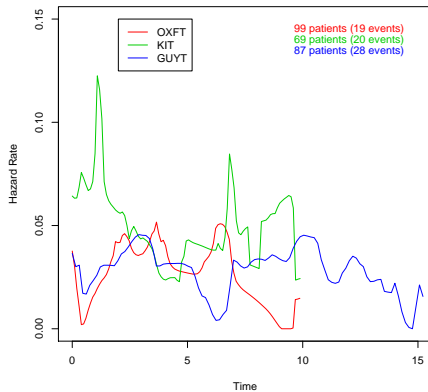
- Biological question : " Can we predict which patients will resist to the Tamoxifen treatment in an adjuvant setting ?"
- Tamoxifen treated patients coming from 3 different institutions, can we pool the data ?
 - ▶ gene-expressions : normalization
 - ▶ survival data : model fitting.
- 255 eligible patients (samples)
- 44928 probes (variables)

Survival Data

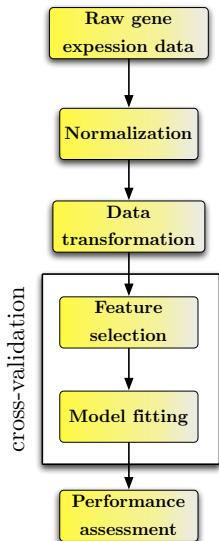
Tamoxifen treated patients
survival curves



Tamoxifen treated patients
hazard functions



Tamoxifen Analysis



In order to keep the design simple, we fix :

- the number of models to aggregate (see feature selection step)
- the cutoff selection (see the model fitting step).

Normalization

- Goal : reduce the systematic variability between samples.
- Method : normalization.

Procedure

- 1 Background correction, expression quantification and normalization were performed using Robust Multichip Average [Irizarry et al., 2003, Bolstad et al., 2003].
- 2 RMA performed separately per population
- 3 Gene median centering per population.

➔ No artifact highlighted by unsupervised clustering.

Data Transformation

- Goal : reduce the dimensionality of the problem.
- Method : cluster highly correlated variables.

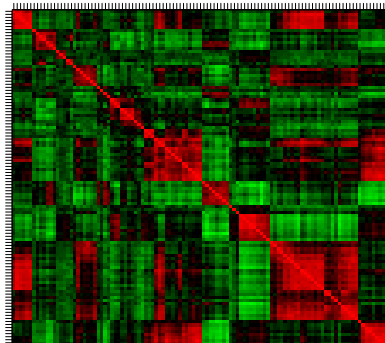
Procedure

- 1 Use of an independent dataset of untreated patients (137 patients, 44929 probes)
- 2 Filtering of the less variant probes.
- 3 Hierarchical clustering (average linkage, uncentered Pearson correlation).
- 4 Cut the tree at height 0.5.
- 5 Keep only clusters with at least 5 known UniGenes.

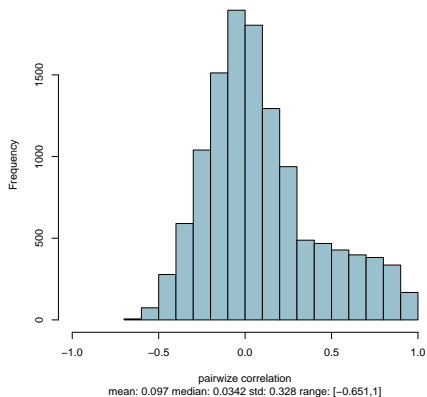
➡ 110 clusters where half are significantly associated with survival (on the Tamoxifen dataset).

Data Transformation

Feature Pairwise Correlation



Histogram of pairwise correlations



- Goal : fast selection of the relevant features.
- Method : ranking based on univariate model significance.

Procedure

- 1 For each feature, compute the likelihood ratio test of the univariate Cox model.
- 2 Perform the ranking based on the p-value.
- 3 Select the top n features (n is fixed).

➡ n features are selected.

- Goal : fit a simple model with low variance.
- Method : aggregation of n univariate classifiers.

Procedure

- 1 The univariate models for the n top features were computed during the previous step.
- 2 Compute a linear combination of these classifiers with weight of 1.

➔ Continuous score representing the risk of a patient.

- We do binary classification from survival data.
- We can not use traditional statistics (sensitivity, specificity, χ^2 test, ...)

➡ Adaptation of such estimators to deal with censoring.

Survival Statistics for 2 Groups

There exist several ways to assess difference in survival between 2 groups

- **Kaplan-Meier estimator** and **Logrank test** :

- ▶ KM method estimates survivor function such that

$$\widehat{S}(t) = \prod_{j:t_j \leq t} \left[1 - \frac{\overbrace{d_j}}{\underbrace{n_j}} \right].$$

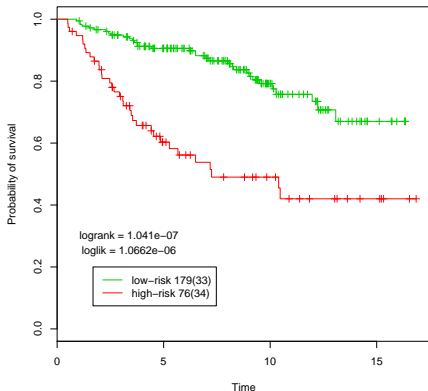
- ▶ logrank method tests $H_0 : S_1(t) = S_2(t) \forall t \geq 0$.

- **Hazard ratio** (HR) : relative hazard between 2 groups using Cox model with one dummy variable ($G = 0/1$ for low and high-risk groups).
- **Time-dependent ROC Curves** and area under ROC curves : extension of traditional ROC curves dealing with censoring data.

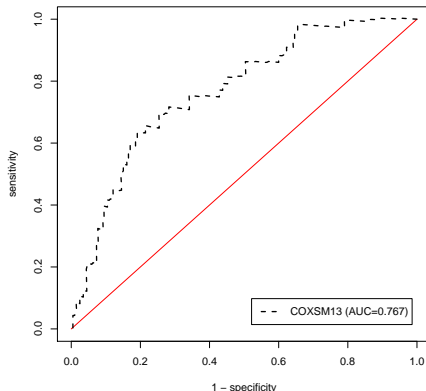
Performance in LOO

Logrank and Time-Dependent ROC Curve

Survival curves
COXSM13 (LOO classification)



Tamoxifen treated patients
time-dependent ROC curves (5 yrs)



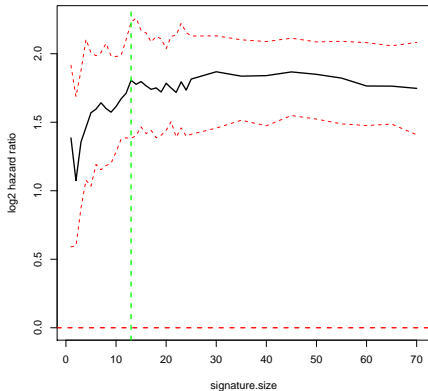
Performance

Hazard Ratio and Logrank

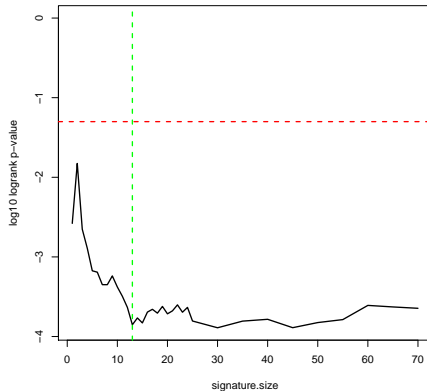
Training set	Test set	Hazard ratio	Log-rank p
OXFT (99/19)	KIT/GUYT (156/48)	2.17 [1.2,3.91]	8.46e-3
KIT (69/20)	OXFT/GUYT (186/47)	4.07 [2.23,7.41]	7.98e-7
GUYT (87/28)	OXFT/KIT (168/39)	5.93 [3,11.75]	1.24e-9
KIT/GUYT (156/48)	OXFT (99/19)	14.59 [5.38,39.52]	1.74e-11
OXFT/GUYT (186/47)	KIT (69/20)	3.44 [1.36,8.67]	5.27e-3
OXFT/KIT (168/39)	GUYT (87/28)	2.23 [1.05,4.71]	3.11e-2
Leave-one-out c.v.		3.85 [2.32;6.41]	1.04e-7
Multiple 10-fold c.v.		3.28 [2.66,3.84]	9.04e-7

Performance wrt Signature Size

Hazard ratio (CI) wrt signature size
10FOLD CV



Logrank p-value wrt signature size
10FOLD CV

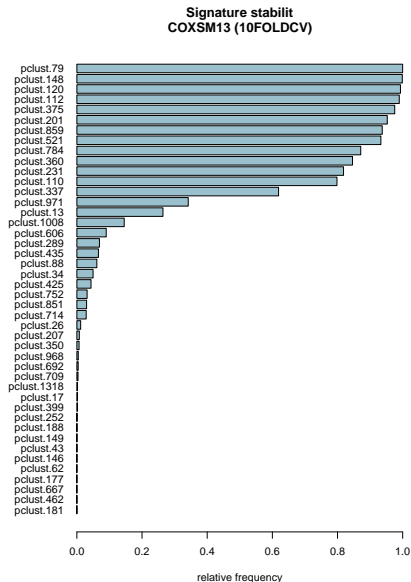


Performance vs Random

- At level of performance estimation :
 - ▶ 1000 random permutations of the labels and perform the whole procedure
 - ➔ only 1% of such classifications gives better discrimination.
- At level of feature selection :
 - ▶ random selection of n features
 - ➔ most of the feature selection result in good classifiers because of the high proportion of relevant features.

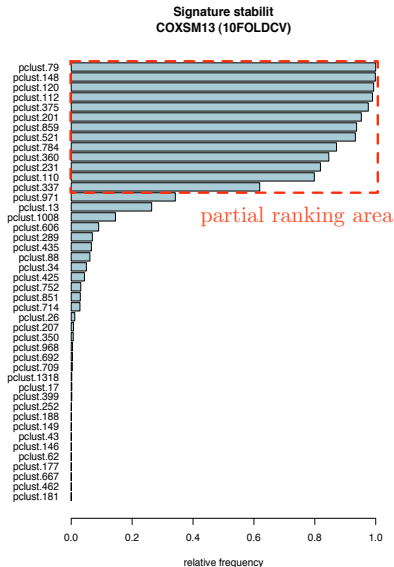
 - ▶ use of the "anti"-ranking
 - ➔ very poor performance.

Feature Selection Stability



- Over the multiple 10-fold cross-validations, several top n features are selected
- If the same set of features are selected every time, we see high relative frequency for these features.

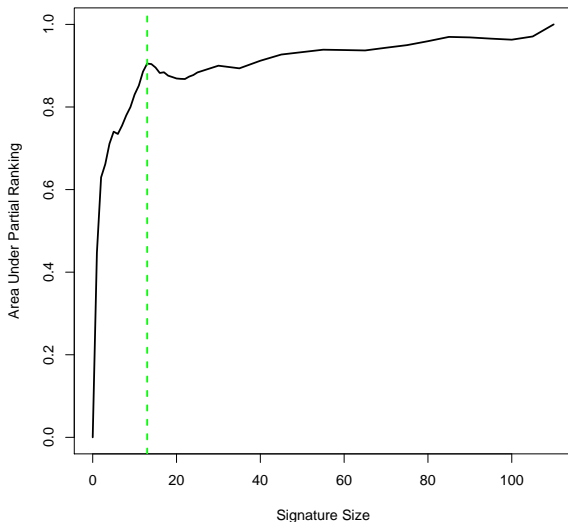
Feature Selection Stability



- Over the multiple 10-fold cross-validations, several top n features are selected
 - If the same set of features are selected every time, we see high relative frequency for these features.
- ➔ We can compute the *area under partial ranking* w.r.t. the number of selected features.

Stability wrt Signature Size

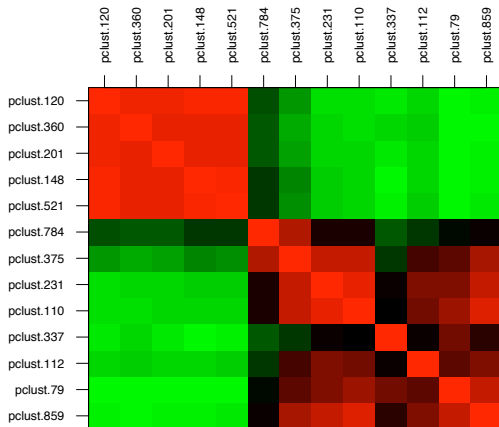
Partial ranking stability wrt signature size
10FOLD CV



- $n = 13$ seems to be a good trade-off between the number of selected features (signature) and its stability.

Final Model

- Use the same method with all the samples.
- The result is a model with a set of features.
- The model and the features are expected from previous results.



- Study the stability of the initial clustering (data transformation step).
- Use of Gene Ontology to study the signature in a biological point of view.
- Objective comparison with the traditional clinical variables.

Current Research Interests

- Meta-analysis.
- Ranking statistics.
- Input space transformation (using biological knowledge, such that gene list enrichment or GO).
- Optimization framework for binary classification of survival data.

Thank you for your attention.



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A comparison of normalization methods for high density
oligonucleotide array data based on variance and bias.
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Irizarry, R. A., Boldstad, B. M., Collin, F., Cope, L. M., Hobbs, B.,
and Speed, T. R. (2003).
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Nucleic Acids Research, 31(4).